



One-pot derivatization of medicinally important 9-aminoacridines by reductive amination and S_NAr reaction

Gary Gellerman*, Vladimir Gaisin, Tamara Brider

Department of Biological Chemistry, Ariel University Center of Samaria, PO Box 3, Ariel 40700, Israel

ARTICLE INFO

Article history:

Received 15 October 2009

Revised 25 November 2009

Accepted 4 December 2009

Available online 11 December 2009

Keywords:

9-Aminoacridine
Reductive alkylation
Nucleophilic substitution
One-pot synthesis

ABSTRACT

A new highly efficient one-pot derivatization of medicinally important 9-aminoacridines (9-AA) at the amine position is described. Simple reductive amination and S_NAr reaction using easily accessible starting materials give a fast entry to novel 9-AA derivatives for biological screening.

© 2009 Elsevier Ltd. All rights reserved.

The 9-aminoacridine (9-AA) core is a structure of interest to medicinal chemists and appears in many biologically active compounds, mostly with anticancer and antimalarial applications. 9-AA derivatives such as quinacrine¹ are able to intercalate with DNA, and consequently, can inhibit DNA transcription in parasites.² 9-Anilinoacridines have good antimalarial activities and are potent parasite DNA topoisomerase II inhibitors.³ N-alkylated 9-AA analogs have been shown to be potent inhibitors of prion disease in cultured neuroblastoma cells, which also show inhibition by lysosomotropic agents and cysteine protease inhibitors.⁴

In the field of antitumor DNA-intercalating agents, 9-AA derivatives play an important role due to their antiproliferative properties.⁵ Several cancer chemotherapeutics such as Amascrine and Ledakrin based on the 9-aminoacridine scaffold have been developed.^{6a} In addition, a series of potential topoisomerase II-mediated anticancer 9-anilinoacridines, which are designed to avoid bio-oxidation and which possess long durations of drug action, have been reported.⁷ Among these substances, 3-(9-acridinylamino)-5-hydroxymethylaniline (AHMA) (Fig. 1) and its alkylcarbamate derivatives have been developed for potential clinical application.⁸

9-Aminoacridines have also been investigated as potential photoaffinity labels⁹ and as fluorescent probes used to detect cancer cells.¹⁰

Recently, 9-aminoacridine analogs, including the antimalarial drug quinacrine, were found to present strong induction of p53 function in renal cell carcinomas (RCCs) and other types of cancer cells.¹¹ Interestingly, induction of p53 function by these com-

pounds does not involve genotoxic stress and is mediated by the suppression of NF- κ B activity. Active NF- κ B signaling provides selective advantages to tumor cells by inhibiting apoptosis and promoting proliferation by stimulating expression of antiapoptotic factors.

These findings, together with those mentioned above, indicate that 9-AA derivatives have potential for anticancer applications and this has inspired us to look for a short and efficient method for the derivatization of 9-AA suitable for the rapid generation of new compounds for evaluation.

Herein we report a new, highly-efficient, one-pot derivatization of 9-AA at the amine position by simple reductive amination or S_NAr reaction, using commercially available synthons. Reductive amination of aldehydes and ketones is an important and direct method for the transformation of these functionalities into amines.¹² In this carbon–nitrogen bond-forming process, the intermediate imine is not pre-formed.¹³ Thus, the chosen reducing agent should be stable enough for the in situ formation of the imine and avoid undesirable carbonyl reduction to alcohol by-products. Mild NaCNBH₃ in a weak acidic media is a widely reported reducing reagent for reductive amination and was therefore employed as the reagent of choice. To demonstrate the synthetic potential of NaCNBH₃ in the reductive amination of 9-AA under standard conditions (1% AcOH in MeOH, rt, 3 h) we employed two classes of aldehydes: aromatic and aliphatic (Scheme 1), yielding *N*(9)-benzylacridines **1a–h** and 2-(acridin-9-ylamino)acetic acid **1i**, respectively, in moderate to good yields (Table 1).¹⁴ The *N*(9)-aminobenzyl analogs synthesized bear representative electron-donating (ED) and electron-withdrawing (EW) groups as well as aryl and indolyl systems at various positions around the benzyl

* Corresponding author. Tel.: +972 3 937 1442; fax: +972 3 906 6634.
E-mail address: garyg@ariel.ac.il (G. Gellerman).

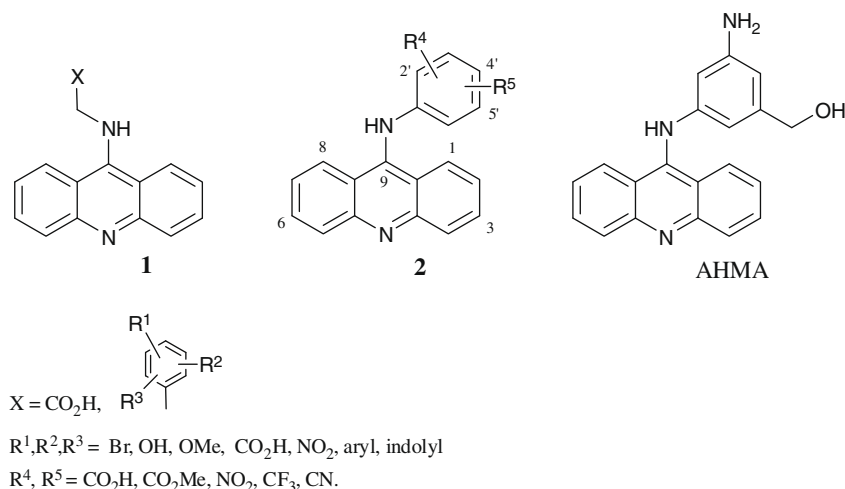
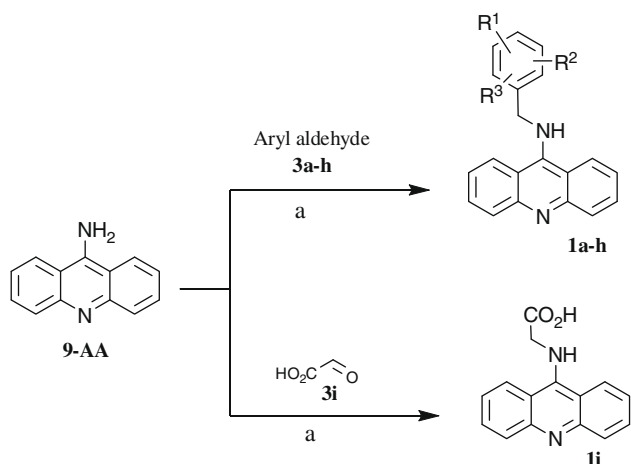


Figure 1. General structures of 9-aminoacridines.



Scheme 1. Synthesis of 9-aminoacridine derivatives **1** by reductive amination. Reagents and conditions: (a) NaCNBH_3 , 1% AcOH in MeOH , 3 h, rt.

ring. Such syntheses are more difficult to accomplish using the classical 'reverse' synthetic approach in which 9-chloroacridine reacts with benzylamines,¹⁵ due to the poor commercial availability of substituted benzylamines. The aliphatic analog **1i** was prepared in the same manner using the corresponding glyoxylic acid hydrate (**3i**).

In the context of exploring the rapid derivatization of the 9-AA scaffold we found that the amine (NH_2) at position nine is nucleophilic enough to take part in nucleophilic aromatic substitution ($\text{S}_{\text{N}}\text{Ar}$) to give medicinally important 9-anilinoacridines.¹⁶

We successfully reacted representative electrophilic haloaryls **4a–e** (Scheme 2), bearing one or two strongly electron-withdrawing groups, with 9-AA in the presence of one equivalent (half molar ratio) of Cs_2CO_3 in DMF at 90 °C to afford the substituted 9-anilinoacridines **2a–e** in good yields (Table 2).¹⁷ The uniqueness of this method is in its ability to form the anilino tether in 9-AAs with two EW groups which is very difficult to accomplish using the standard 'reverse' approach, namely nucleophilic substitution of the deactivated anilines on 9-chloroacridines.

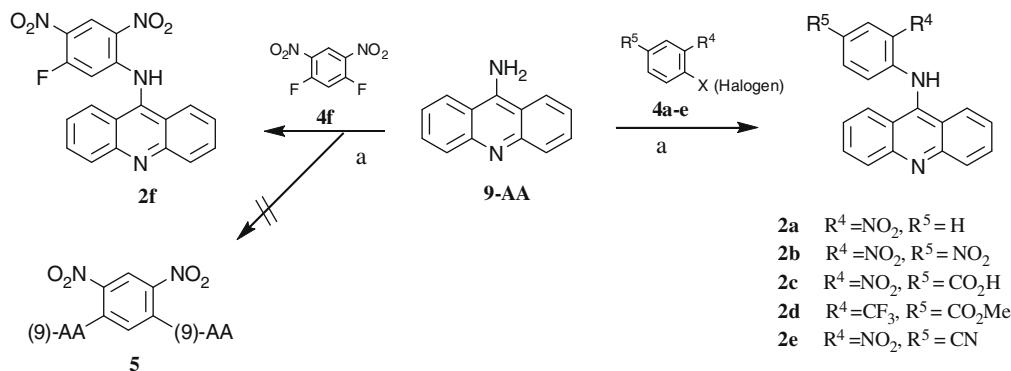
The anilino amine in such a 'reverse' reaction is strongly deactivated by EW groups leading mostly to unreacted materials or black tars. The additional advantage of the $\text{S}_{\text{N}}\text{Ar}$ reaction with 9-AAs is the extensive commercial availability of appropriately substituted haloaryls. Interestingly, 4-chloro-3-nitrobenzoic acid

Table 1
Reaction data for reductive amination of 9-AA

Entry	Aldehyde	Product	Yield (%)
1	3a 	1a	92
2	3b 	1b	89
3	3c 	1c	87
4	3d 	1d	66 ^a
5	3e 	1e	68 ^a
6	3f 	1f	72 ^a
7	3g 	1g	83
8	3h 	1h	58 ^a
9	3i 	1i	91

^a After chromatography.

(**4c**) which bears an acidic CO_2H group smoothly undergoes the $\text{S}_{\text{N}}\text{Ar}$ reaction to give product **2c** even in the presence of basic Cs_2CO_3 . Moreover, the CO_2H group in **2c** (as well as in the



Scheme 2. Synthesis of 9-aminoacridine derivatives **2** by S_NAr reaction. Reagents and conditions: (a) Cs₂CO₃, DMF, 90 °C, 12 h.

Table 2
Reaction data for S_NAr reaction of 9-AA

Entry	Haloarene	Product	Yield (%)
1		2a ¹⁸	73 ^a
2		2b	78 ^a
3		2c	86
4		2d	76 ^a
5		2e	71 ^a
6		2f	52 ^a

^a After chromatography.

previously mentioned carboxy and hydroxy analogs **1a–c, g, and i**) could serve as a linking group to various carriers for possible delivery. Another attempt was made to obtain bis-9-AA **5** (Scheme 2) by reacting 2.5 equiv of 9-AA with 1 equiv of 1,5-difluoro-2,4-dinitrobenzene. Unfortunately, only the mono adduct **2f** was obtained in a moderate yield, most probably due to severe steric hindrance.

All the products synthesized in this work which possess acidic CO₂H or phenolic substitution (**1a, b, c, g, i, and 2c**) precipitated from acetone as pure (more than 94% by HPLC) yellow solids, while unreacted starting materials and solvents were completely soluble in acetone. Such a phenomenon can be attributed to the formation of poorly soluble (in organic solvents) zwitterions. This hypothesis is supported by an observed low-field shift in the ¹H NMR spectrum for the anilinic H-2' in **2c**, *ortho* to the EW positively charged 9-aminoacridinium moiety (δ 7.76, J = 6.5 Hz, see data for **2c** in the Supplementary data). Typically, protons *ortho* to strongly electron-

donating (ED) free amine bases, as in **2b** and **2d**, are shifted higher-field to δ 7.01 and δ 6.85, respectively (see Supplementary data for **2b, d**). Other one-pot derivatizations of 9-AA are under investigation.

In conclusion, we have developed a new method for the efficient one-pot derivatization of the medically important 9-aminoacridine (9-AA) scaffold. A simple reductive amination of 9-AA with aldehydes yielded a series of novel substituted *N*(9)-benzylaminoacridines and *N*(9)-alkylaminoacridines. An S_NAr reaction between 9-AA and halobenzenes that had strong EW groups gave novel *N*(9)-anilinoacridines. All the products were obtained from commercially available synthons in good yields. These synthetic routes provide easy and rapid access to novel 9-AA derivatives which can be further explored for their biological properties.

Acknowledgment

The authors thank Dr. Alexandra Massarwa for the HRMS measurements of all new compounds.

Supplementary data

Supplementary data (selected ¹H and ¹³C NMR, HRMS spectra) associated with this article can be found, in the online version, at doi:10.1016/j.tetlet.2009.12.020.

References and notes

- (a) Chavalitshewinkoon, P.; Wilairat, P.; Ralph, R. *Antimicrob. Agents Chemother.* **1993**, *37*, 403–406; (b) Elueze, E. I.; Croft, S. L.; Warhurst, D. C. J. *Antimicrob. Chemother.* **1996**, *37*, 511–518; (c) Figgitt, D.; Denny, W.; Ralph, R. *Antimicrob. Agents Chemother.* **1992**, *36*, 1644–1647; (d) Shibnev, V. A.; Finogenova, M. P.; Allahverdiev, A. M. *Bioorg. Khim.* **1988**, *14*, 1565–1569; (e) Wainwright, M. J. *Antimicrob. Chemother.* **2001**, *47*, 1–13.
- (a) Hahn, F. E.; Fean, C. L. *Antimicrob. Agents Chemother.* **1969**, *9*, 63–66; (b) Allison, J. L.; O'Brien, R. L.; Hahn, F. E. *Antimicrob. Agents Chemother.* **1965**, *5*, 310–314; (c) Hahn, F. E. *Antibiotics* **1974**, *3*, 58–78.
- (a) Gamage, S. A.; Tepsiri, N.; Wilairat, P.; Denny, W. A. *J. Med. Chem.* **1994**, *37*, 1486–1494; (b) Auparakkitanon, S.; Wilairat, P. *Biochem. Biophys. Res. Commun.* **2000**, *269*, 406–409.
- Doh-ura, K.; Iwaki, T.; Caughey, B. J. *Virology* **2000**, *74*, 4894–4897.
- Sebestik, J.; Hlavacek, J.; Stibor, I. *Curr. Protein Pept. Sci.* **2007**, *8*, 471–483.
- (a) Denny, W. A. *Curr. Med. Chem.* **2002**, *9*, 1655–1665; (b) Anderson, B.; Sherrill, J.; Kiplin, G. R. *Bioorg. Med. Chem.* **2006**, *14*, 334–343.
- (a) Jaycox, G. D.; Gribble, G. W.; Hacker, M. P. *J. Heterocycl. Chem.* **1987**, *24*, 1405–1408; (b) Ciesielska, E.; Pastwa, E.; Szmigiero, L. *Acta Biochim. Pol.* **1997**, *44*, 775–780; (c) Walker, T. M.; Starr, B.; Atterwill, C. *Human Exp. Toxicol.* **1995**, *14*, 469–474; (d) Bonse, S.; Santelli-Rouvier, C.; Krauth-Siegel, R. L. *J. Med. Chem.* **1999**, *42*, 5448–5454; (e) Inhoff, O.; Richards, J. M.; Krauth-Siegel, R. L. *J. Med. Chem.* **2002**, *45*, 4524–4530; (f) Obexer, W.; Schmid, C.; Brun, R. *Trop. Med. Parasitol.* **1995**, *46*, 49–53.
- Kopacz, S. J.; Mueller, D. M.; Lee, C. P. *Biochim. Biophys. Acta* **1985**, *807*, 177–188.
- Schwarz, G.; Wittekind, D. *Anal. Qual. Cytol.* **1982**, *4*, 44–54.
- Kapuriya, N.; Kapuriya, K.; Tsann-Long, S. *Bioorg. Med. Chem.* **2008**, *16*, 5413–5423.

11. Gurova, K. V.; Hill, J. E.; Gudkov, A. V. *PNAS* **2005**, *102*, 17448–17453.
12. Hutchins, R. O.; Hutchins, M. K. In *Comprehensive Organic Synthesis*; Trost, B. N., Ed.; Pergamon: Oxford, 1991; Vol. 8, p 25.
13. Burkhardt, E. R.; Coleridge, B. M. *Tetrahedron Lett.* **2008**, *49*, 5152–5155.
14. *General procedure for the synthesis of 1a–i via reductive amination*: 9-AA (0.194 g, 1 mmol) and the corresponding aldehyde (1 mmol) were added to 5 mL of MeOH/AcOH (99:1) and the mixture stirred at room temperature until the starting material dissolved (15 min). Then, NaCNBH₃ (0.09 g, 1.5 mmol) was added in small portions with stirring. After additional stirring for 3 h at room temperature, the solvent was evaporated and the residue was taken into acetone. The resulting precipitate was filtered under vacuum, washed with acetone, and dried to give the product as a yellow solid. Compounds that did not precipitate were purified by flash column chromatography on silica gel 60 (5% MeOH in EtOAc) to yield pure (yellow) products. Data for **1a** (0.3 g, 92% yield): ν_{\max} (KBr): 3500–3180 (br, s), 1700 (C=O), 1640, 1290 cm⁻¹; HRMS (Cl, *m/z*) calcd for C₂₁H₁₇N₂O₂ (MH⁺) 329.121, found 329.102; ¹H NMR (300 MHz, DMSO-*d*₆): δ 8.50 (d, 2H, *J* = 7.0 Hz), 7.92 (d, 2H, *J* = 7.0 Hz), 7.77 (d, 1H, *J* = 6.8 Hz), 7.68–7.65 (m, 2H), 7.41–7.36 (m, 2H), 7.30–7.24 (m, 3H), 4.54 (s, 2H, –NH–CH₂–); ¹³C NMR (75 MHz, CDCl₃): 171.9 (CO₂H), 168.7, 152.7, 146.5, 141.5, 139.0, 131.5, 130.6, 130.1, 128.6, 128.2, 126.3, 125.5, 123.6, 122.1, 64.1.
15. Guetzoyan, L.; Ramiandrasoa, F.; Perree-Fauvet, M. *Bioorg. Med. Chem.* **2007**, *15*, 3278–3289.
16. *Advanced Organic Chemistry, Reactions, Mechanisms and Structure*, Jerry March 3rd edition.
17. *General procedure for the synthesis of 2a–e via S_NAr reaction*: 9-AA (0.194 g, 1 mmol), haloaryl compound (1 mmol), and Cs₂CO₃ (0.161 g, 0.5 mmol) were heated in 5 mL of dry DMF at 90 °C for 12 h. While heating, the color of the reaction mixture changed to dark red. After completion of the reaction (TLC monitoring in CH₂Cl₂) the mixture was cooled and poured into water. In the case of **2c**, the pH was adjusted to 6 by careful addition of 0.1 N HCl. The resulting precipitate was collected by filtration, washed several times with water, and dried to give a crude orange solid. The products (except **2c** which precipitated in pure form) were purified by flash column chromatography on silica gel 60 (CH₂Cl₂). Data for **2a** (orange solid, 0.23 g, 73% yield): ν_{\max} (KBr): 1670, 1650, 1190 cm⁻¹; HRMS (Cl, *m/z*) calcd for C₁₉H₁₄N₃O₂ (MH⁺) 316.101, found 316.061; ¹H NMR (300 MHz, CDCl₃): δ 9.32 (br s, 1, NH), 8.24 (d, 2H, *J* = 6.8 Hz), 8.01–7.88 (m, 3H), 7.71–7.53 (m, 4H), 7.33 (t, 1H, *J* = 7.0 Hz), 6.62 (d, 1H, *J* = 7.0 Hz). ¹³C NMR (75 MHz, CDCl₃): 165.2, 154.3, 147.7, 144.1, 133.4, 130.0, 129.7, 128.7, 127.2, 125.6, 124.7, 122.3, 120.5.
18. Denny, W. A.; Cain, B. F. *J. Med. Chem.* **1982**, *25*, 276–315.